

¹⁵N-CIDNP investigations during tryptophan, N-acetyl-L-tryptophan, and melatonin nitration with reactive nitrogen species

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Abstract

Tryptophan and melatonin are nitrated by peroxynitrite; tryptophan residues in proteins are susceptible to attack by reactive nitrogen species. Nitrated tryptophan might therefore be used as a biomarker for the involvement of reactive species derived from nitrogen oxide in a variety of pathophysiological conditions. The radical character of the tryptophan (Trp) and N-acetyl-L-tryptophan (N-AcTrp) nitration with peroxynitrite is shown using ¹⁵N-CIDNP. During the decay of peroxynitrite-¹⁵N in the presence of Trp at pH 5 in the probe of a ¹⁵N-NMR spectrometer, the ¹⁵N-NMR signals of various nitrated tryptophans (¹⁵NO₂-Trp) show emission (E). The effects are built up in radical pairs [Trp, ¹⁵NO₂]^F formed by diffusive encounters of radicals ¹⁵NO₂ and Trp generated during decay of peroxynitrite-¹⁵N in the presence of Trp. Similar ¹⁵N-CIDNP effects are observed during reaction of Trp and/or N-AcTrp using the nitrating systems H¹⁵NO₃, H¹⁵NO₄ and H₂O₂/¹⁵NO₂⁻/HRP, which are also built up in radical pairs [Trp, ¹⁵NO₂]^F. During nitration of melatonin (Mel) with peroxynitrite-¹⁵N and H¹⁵NO₄, the ¹⁵N-NMR signal of 4-nitromelatonin (4-¹⁵NO₂-Mel) shows emission arising from radical pairs [Mel, ¹⁵NO₂]^F which are formed in an analogous manner.

Keywords: Tryptophan, 6-nitrotryptophan, melatonin, peroxynitrite, reactive nitrogen species (RNS), nitrogen dioxide

Abbreviations: 1, 3-methylindane; 2, 3-methyl-5-methoxyindane; A, enhanced absorption; AcOH, acetic acid; c, cage product; CIDNP, chemically induced dynamic nuclear polarization; E, emission; e, escape product; F, radical precursor; g, g value; HRP, horseradish peroxidase; Mel, melatonin (N-[2-(5-methoxy-1H-indol-3-yl)ethyl]-acetamide; MetMb, Metmyoglobin; MPO, myeloperoxidase; N, no CIDNP effect; N-AcTrp, N-acetyl-L-tryptophan; N-AcTrp-NH₂, N-acetyl-L-tryptophan amide; NO₂-Mel, nitromelatonin; NO₂-N-AcTrp, nitro-N-acetyl-L-tryptophan; NO₂-Trp, nitrotryptophan; NO₂-Tyr, nitrotyrosine; RNS, reactive nitrogen species; S, singlet state

Introduction

Peroxynitrite (O=NO-OH, pK 6.5–6.8) is known as an unstable intermediate during reaction of hydrogen peroxide (H₂O₂) with nitrous acid (HNO₂) [1]. In recent years, peroxynitrite—this term is commonly used for the acid-base equilibrium mixture of peroxynitrous acid (O=NO-OH) and peroxynitrite

anion (O=NOO⁻)—has received considerable attention as a putative cytotoxic agent in living organisms where it may be produced by recombination of endogenous nitric oxide (NO) and superoxide anion (O₂⁻) [2]. It isomerizes to nitrate or may react with L-tyrosine (Tyr) forming 3-nitrotyrosine (3-NO₂-Tyr) which is a useful marker of oxidants like peroxynitrite [3,4]. It also reacts with L-tryptophan (Trp) and

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used as a well-known nitrating agent in strongly acidic media. The application of HNO_4 to *N*-AcTrp nitration at medium pH will complete this study [15]. In a further part, the nitration of the Trp derivative melatonin (*N*-[2-(5-methoxy-1*H*-indol-3yl)-ethyl]-acetamide; Mel) will be described, too. Mel is recognized as a scavenger of strong oxidants [16].

Materials and methods

^{15}N -CIDNP experiments with preformed and in situ generated peroxyxynitrite- ^{15}N

Stock solutions of peroxyxynitrite- ^{15}N ($0.435 \pm 0.01 \text{ M O}=\text{}^{15}\text{NOO}^-$) were prepared by reaction of *iso*-amylxynitrite- ^{15}N (0.0024 mol) with hydrogen peroxide (2 ml H_2O_2 , 1 M) in diethylene-triaminepentaacetic acid-free solutions and purified (six times solvent extraction with *n*-hexane, removal of excess H_2O_2 by passing over MnO_2 , N_2 -purging) [17], divided into 200- μl aliquots in 1-ml Eppendorf vials and stored at 194 K in the dark. The reaction mixtures were prepared in 10-mm NMR tubes by adding peroxyxynitrite- ^{15}N to the frozen solvent ($\text{H}_2\text{O}/\text{D}_2\text{O}$: 9/1) containing phosphate buffer (0.3 M), if required, NaHCO_3 (0.05 M) and Trp or *N*-AcTrp (0.05 M) at 268 K. The final pH after mixing was 7.5. The tubes were quickly transferred into the probe of the ^{15}N -NMR spectrometer (Bruker DPX-300) and locked (internal lock: D_2O). One minute after mixing of the reactants, the first ^{15}N -NMR spectrum was taken by using single pulses with pulse angles of 90° . This procedure was repeated every minute until the reaction was completed. For detecting reaction products, ^{15}N -NMR spectra were taken with several pulses at room temperature.

For generating peroxyxynitrite- ^{15}N *in situ*, a solution of $\text{Na}^{15}\text{NO}_2$ (0.05 M) in $\text{H}_2\text{O}/\text{D}_2\text{O}$ (9:1) containing phosphate buffer (0.3 M) and, when needed, NaHCO_3 (0.05 M) and Trp or *N*-AcTrp (0.05 M) was prepared in 10 mL tubes at pH 5, and a single ^{15}N -NMR spectrum was taken using a 90° pulse. After that, the tube was replaced, and H_2O_2 (1 M) was added to the solution. ^{15}N -NMR spectra were then taken every 1–5 min until the reaction was completed.

^{15}N -CIDNP experiments with the systems $\text{H}_2\text{O}_2/^{15}\text{NO}_2^-/\text{HRP}$, H^{15}NO_3 , and H^{15}NO_4

Solutions of HRP (20 μM), H_2O_2 (1 M), $\text{Na}^{15}\text{NO}_2$ (0.05 M) and *N*-AcTrp were prepared at pH 7 [18]. Spectra were taken as described above. Nitration of *N*-AcTrp (0.05 M) with H^{15}NO_3 was performed by adding *N*-AcTrp to a solution of H^{15}NO_3 (0.5 M) in AcOH with 10% D_2O [19]. H^{15}NO_4 was prepared *in situ* by adding H_2O_2 (1 M) to a solution of $\text{Na}^{15}\text{NO}_2$ (0.15 M) at pH 2 [20]. After about 2 min, *N*-AcTrp (0.05 M) was added [15].

Materials and solutions

It was taken care to exclude possible contamination by CO_2 and transition metal ions. The following procedures were performed under nitrogen and at reduced incident light. Doubly distilled water was bubbled (21 min^{-1}) with nitrogen (5.0) at room temperature for 20 min and was then treated with the heavy metal ion scavenger resin Chelex 100 (0.5 g in 10 ml) by gently shaking for 18 h in the dark. After separation from the resin by low-speed centrifugation for 5 min and careful decanting, the water was again bubbled with nitrogen for 20 min.

$\text{Na}^{15}\text{NO}_2$ and *iso*-amylxynitrite- ^{15}N were purchased from Aldrich/Isotec Inc. (Taufkirchen, Germany). HNO_3 was 9.4 M in H_2O and labelled with 60.3 atom% ^{15}N . All other chemicals were from Sigma (Deisenhofen, Germany) and were of the highest purity available.

Calculation of ^{15}N chemical shifts

Isotropic absolute shielding constants were calculated using the GIAO protocol at the DFT/aug-cc-pVDZ/DFT/aug-cc-pVDZ level of theory. During these calculations solvation corrections (CH_3CN for nitrobenzol, H_2O for all others) with the CPCM solvation model were performed at the same level of theory [21].

Results and discussion

^{15}N -CIDNP during nitration of *N*-AcTrp with H^{15}NO_3 in AcOH

Nitration of Trp and its derivatives *N*-AcTrp and *N*-AcTrp- NH_2 with RNS has thoroughly been studied [6,22,23], see Table I. It mainly occurs at the 6-position of the indole system, leading to 6- NO_2 -Trp, 6- NO_2 -*N*-AcTrp, and 6- NO_2 -*N*-AcTrp- NH_2 . Besides, nitration at the 1-, 4-, 5- and 7- positions are reported (Figure 1). Aromatic compounds react with nitric acid in a non-radical way, if toluene and less activated compounds are used. Electron rich aromatic systems are nitrated via radicals in a nitrous acid catalysed path which can be proved using ^{15}N -CIDNP [14,26]. Indole derivatives are nitrated at various ring positions [27]. During reaction of *N*-AcTrp- NH_2 in AcOH, nitration at the 4-, 6- and 7-position are reported [25]. The nitration mechanism of indoles with nitric acid has not yet been studied.

During reaction of H^{15}NO_3 with *N*-AcTrp in AcOH, a ^{15}N -NMR spectrum was taken which is shown in Figure 2(A), details of the reaction are given in Table II. Besides the ^{15}N -NMR signal at $\delta = -15$ ppm due to H^{15}NO_3 , emission signals at $\delta = -10, 2, 4, 18$ and 19 ppm are observed which are caused by nitration products of *N*-AcTrp. After reaction, a spectrum was taken with 167 scans and is given in Figure 2(B).

Table I. Nitration products of Trp and its derivatives with RNS [6]*.

System	Trp	Nitrated products	Ref.
O=NOO ⁻	Trp	6-NO ₂ -Trp 5-NO ₂ -Trp 4- or 7-NO ₂ -Trp (minor)	[22]
O=NOO ⁻ (0.5 mM) (5–10 mM)	Trp	6-NO ₂ -Trp	[23]
O=NOO ⁻	Trp N-AcTrp	6-NO ₂ -Trp (major) + other NO ₂ -Trps 1-NO ₂ -N-AcTrp 6-NO ₂ -N-AcTrp	[5]
H ₂ O ₂ /NO ₂ ⁻ /MPO	N-AcTrp	1-NO ₂ -N-AcTrp 6-NO ₂ -N-AcTrp	[5]
H ₂ O ₂ /NO ₂ ⁻ /MetMb	Trp	4-NO ₂ -Trp 5-NO ₂ -Trp 6-NO ₂ -Trp	[24]
H ₂ O ₂ /NO ₂ ⁻ /HRP or HNO ₃ in AcOH	N-AcTrp-NH ₂	1-NO ₂ -N-AcTrp-NH ₂ [†] 4-NO ₂ -N-AcTrp-NH ₂ 6-NO ₂ -N-AcTrp-NH ₂ 7-NO ₂ -N-AcTrp-NH ₂	[25]
O=NOO ⁻	Mel	4-NO ₂ -Mel 6-NO ₂ -Mel	[16]

* MPO, myeloperoxidase; N-AcTrp-NH₂, N-acetyl-L-tryptophan amide; MetMb, metmyoglobin; [†] Not formed with HNO₃.

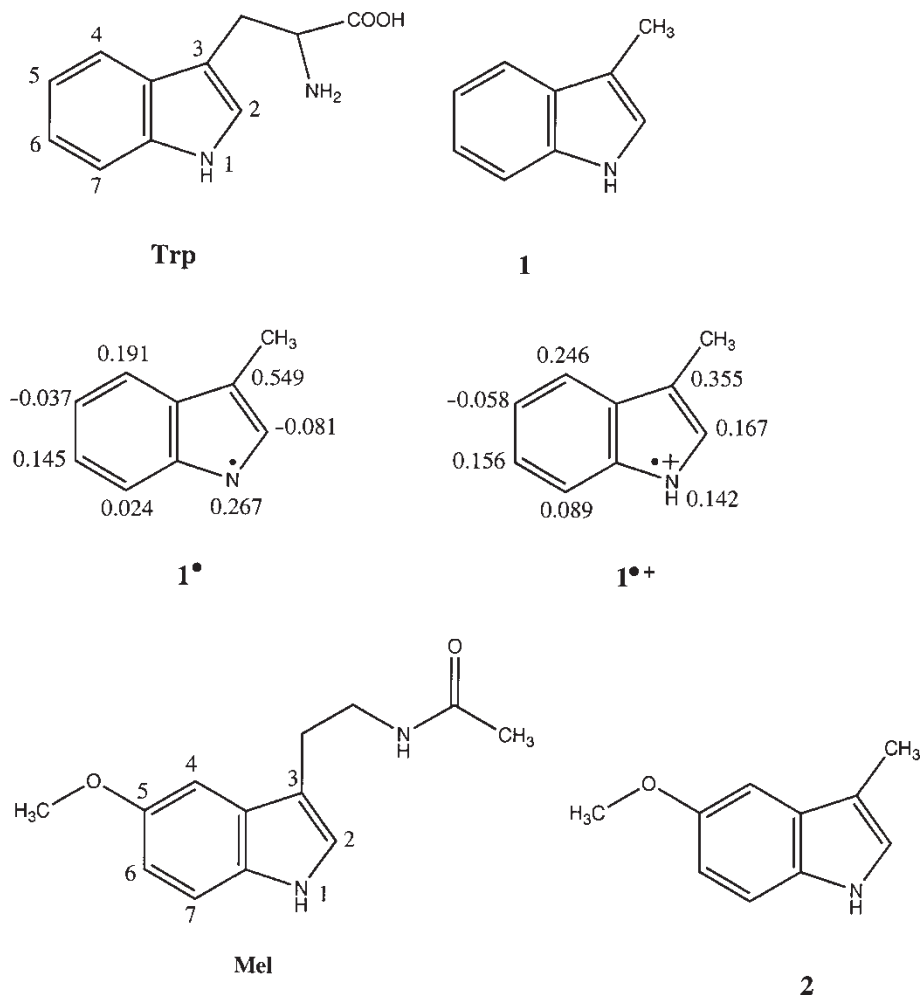


Figure 1. Indane derivatives and spin densities in radicals 1^\bullet and radical cations $1^{\bullet+}$. Trp, Tryptophan; 1, 3-Methylindane; Mel, Melatonin; 2, 3-Methyl-5-methoxyindane.

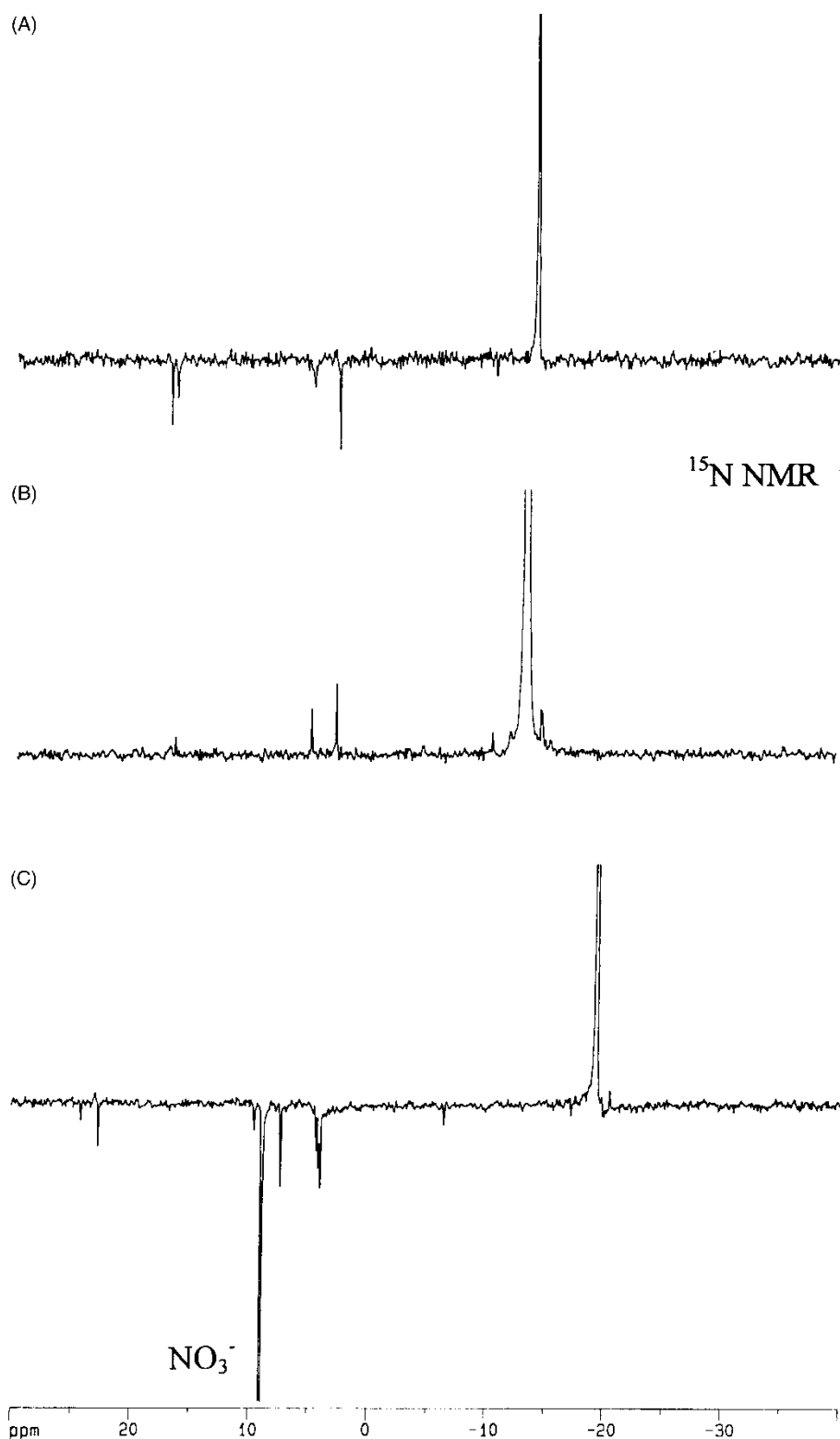


Figure 2. (A) ^{15}N -NMR spectrum during reaction of H^{15}NO_3 with *N*-AcTrp in AcOH/ D_2O 2 min after mixing the reactants (1 scan), (B) 100 min later (167 scans). (C) ^{15}N -NMR spectrum during reaction of H^{15}NO_4 with *N*-AcTrp at pH 2 in $\text{H}_2\text{O}/\text{D}_2\text{O}$ 2 min after mixing the reactants (1 scan).

The signals of the nitration products show weak absorption signals indicating the occurrence of ^{15}N -CIDNP during reaction. Reactions leading to the ^{15}N -CIDNP effects are given in Scheme 3. Analogous effects have been observed during reaction of H^{15}NO_3

with electron-rich aromatics, especially with tyrosine and *N*-acetyl-L-tyrosine (*N*-AcTyr) [9,19], which have similar redox potentials (Trp: 1.015 V, Tyr: 0.93 V at pH 7 [28]), and have been explained in an analogous manner. The effects are built up in radical pairs

Table II. ^{15}N -CIDNP during nitration of *N*-AcTrp with H^{15}NO_3 and H^{15}NO_4 at 298 K.

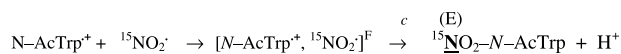
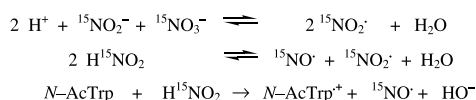
System	^{15}N -NMR signals	CIDNP*
H^{15}NO_3 (0.5 M), <i>N</i> -AcTrp (0.05 M), AcOH, Ph < 1 (Figure 2a)	- 15 (H^{15}NO_3)	N
	- 10 ($2\text{-}^{15}\text{NO}_2\text{-}N\text{-AcTrp}$)	E
	2 ($6\text{-}^{15}\text{NO}_2\text{-}N\text{-AcTrp}$) [†]	E
	4 ($4\text{-}^{15}\text{NO}_2\text{-}N\text{-AcTrp}$)	E
	18 ($3\text{-}^{15}\text{NO}_2\text{-}N\text{-AcTrp}$) [‡]	E
	19 ($3\text{-}^{15}\text{NO}_2\text{-}N\text{-AcTrp}$) [‡]	E
H^{15}NO_4 [¶] , <i>N</i> -AcTrp (0.05 M) pH 2 (Figure 2c)	- 20 (H^{15}NO_4)	A
	- 7 ($2\text{-}^{15}\text{NO}_2\text{-}N\text{-AcTrp}$)	E
	3 ($6\text{-}^{15}\text{NO}_2\text{-}N\text{-AcTrp}$)	E
	7 ($4\text{-}^{15}\text{NO}_2\text{-}N\text{-AcTrp}$)	E
	9 ($^{15}\text{NO}_3^-$)	N

*E: emission, A: enhanced absorption, N: no CIDNP; [†]The signals might also be assigned to 5- and 7- $^{15}\text{NO}_2\text{-}N\text{-AcTrp}$, see Table III;

[‡]Two diastereomeric configurations; [¶]Generated *in situ* by reaction of $\text{Na}^{15}\text{NO}_2$ (0.15 M) with H_2O_2 (1 M).

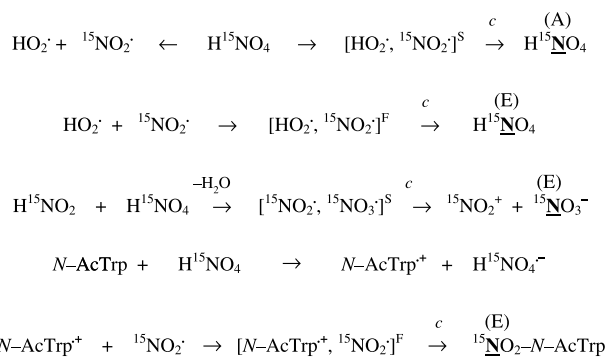
$[N\text{-AcTrp}^{\cdot+}, ^{15}\text{NO}_2]^{\text{F}}$ generated by encounters of radicals $^{15}\text{NO}_2^{\cdot}$ and radical cations *N*-AcTrp^{•+} which are formed independently. Reaction between H^{15}NO_3 and H^{15}NO_2 generates $^{15}\text{NO}_2^{\cdot}$. *N*-AcTrp^{•+} is formed by oxidation with H^{15}NO_2 or, more probably, some reactive intermediate like $^{15}\text{NO}^+$ which is easily formed at strongly acidic conditions.

The emission-type effect in the ^{15}N -NMR signals of $^{15}\text{NO}_2\text{-}N\text{-AcTrp}$ is explained using the observed *g* values of NO_2^{\cdot} (2.000) and Trp^{\cdot} (2.003) [29,30], see Schemes 3 and 4. The reported p*K* value of 4.3 for Trp^{\cdot} indicates that the protonated form of the radical, *N*-AcTrp^{•+}, should be present under the reaction conditions [31]. For assigning the emission signals to distinct nitration products, ^{15}N chemical shifts have been quantum-chemically calculated [21]. For simplicity, 3-methylindane **1** has been chosen as a model compound (Figure 1). The results are listed in Table III. A spin density distribution for the radical cation of **1**, **1**^{•+}, is added, showing that radical recombination reactions are possible at different ring positions [32]. At first, it should be noted that 1- $^{15}\text{NO}_2\text{-}N\text{-AcTrp}$ is obviously not formed, as the corresponding ^{15}N -NMR signal is missing, in agreement with data of Ref. [25]. The small emission signal at $\delta = -10$ ppm might be due to 2- $^{15}\text{NO}_2\text{-}N\text{-AcTrp}$. The strong emission signals at $\delta = 1$ ppm and $\delta = 4$ ppm are assigned to 5-, 6-, 7- $^{15}\text{NO}_2\text{-}N\text{-AcTrp}$ and 4- $^{15}\text{NO}_2\text{-}N\text{-AcTrp}$, respectively, which are the main products, in accordance with the literature, see Table I. The narrow doublet at $\delta = 18$ ppm and $\delta = 19$ ppm is assigned to 3- $^{15}\text{NO}_2\text{-}N\text{-AcTrp}$ which is



Scheme 4. ^{15}N -CIDNP during nitration of *N*-AcTrp with H^{15}NO_3 .

formed in two diastereomeric configurations, in accordance with the formation of cyclohexadienone-like intermediates observed during nitration of tyrosine and related compounds [9,10,14,19].



Scheme 5. ^{15}N -CIDNP during nitration of *N*-AcTrp with H^{15}NO_4 .

Table III. Quantum-chemically calculated isotropic absolute shielding constants and ^{15}N chemical shifts (δ , in ppm against nitrobenzene- ^{15}N).

Molecule	Isotropic shielding constants*		Isotropic chemical shifts	
	B1LYP	B3LYP	B1LYP	B3LYP
Nitrobenzene	-125.2	-121.5	0.0	0.0
NO_3^-	-134.9	-130.2	9.7	8.7
O_2NOOH	-105.6	-105.8	-19.6	-15.7
1- $\text{NO}_2\text{-1}$	-81.7	-77.6	-3.5	-43.9
2- $\text{NO}_2\text{-1}$	-110.7	-106.2	-14.5	-15.3
3- $\text{NO}_2\text{-1}$	-149.0	-149.0	23.8	27.5
4- $\text{NO}_2\text{-1}$	-131.9	-127.7	6.7	6.2
5- $\text{NO}_2\text{-1}$	-127.1	-122.9	1.9	1.4
6- $\text{NO}_2\text{-1}$	-126.9	-122.7	1.7	1.2
7- $\text{NO}_2\text{-1}$	-126.9	-122.6	1.7	1.1

*Isotropic absolute shielding constants were calculated using the GIAO protocol at the DFT/aug-cc-pVDZ//DFT/aug-cc-pVDZ level of theory. During these calculations solvation corrections (CH_3CN for nitrobenzol, H_2O for all others) with the CPCM solvation model were performed at the same level of theory [21].

Table IV. ¹⁵N-CIDNP during nitration of Trp and *N*-AcTrp with peroxyxynitrite-¹⁵N and the system H₂O₂/¹⁵NO₂⁻/HRP at 298 K.

System	¹⁵ N-NMR signals	CIDNP*
Peroxyxynitrite- ¹⁵ N (0.03 M)	8 (¹⁵ NO ₃ ⁻)	N
Trp (0.05 M), 268 K, pH 7.5	242 (¹⁵ NO ₂ ⁻)	N
Peroxyxynitrite- ¹⁵ N [†] , Trp (0.05 M), NaHCO ₃ (0.05 M), pH 5 (Figure 3)	-20 (H ¹⁵ NO ₄) 1 (6- ¹⁵ NO ₂ -Trp) [‡] 4 (4- ¹⁵ NO ₂ -Trp) 8 (¹⁵ NO ₃ ⁻) 15 (3- ¹⁵ NO ₂ -Trp) [¶] 16 (3- ¹⁵ NO ₂ -Trp) [¶] 242 (¹⁵ NO ₂ ⁻) [§]	N E E A E E E
Peroxyxynitrite- ¹⁵ N [†] , <i>N</i> -AcTrp (0.05 M), pH 5 (Figure 4a)	-19 (H ¹⁵ NO ₄) -10 (2- ¹⁵ NO ₂ - <i>N</i> -AcTrp) 0-8 (¹⁵ NO ₂ - <i>N</i> -AcTrp) 8 (¹⁵ NO ₃ ⁻) 242 (¹⁵ NO ₂ ⁻) [§]	E E E N N
HRP (20 μM), H ₂ O ₂ (1 M), Na ¹⁵ NO ₂ (0.05 M), <i>N</i> -AcTrp (0.05 M), pH 7 (Figure 4b,c)	8 (¹⁵ NO ₃ ⁻) 0-8 (¹⁵ NO ₂ - <i>N</i> -AcTrp) 242 (¹⁵ NO ₂ ⁻) [§]	N E N

* E: emission, A: enhanced absorption, N: no CIDNP; [†] Generated *in situ* by reaction of Na¹⁵NO₂ (0.05 M) with H₂O₂ (1 M); [‡] The signals might be assigned to 5- and 7-¹⁵NO₂-Trp, too; [¶] Two diastereoisomeric configurations; [§] Not shown in Figures 3 and 4; ^{||} Assigned to 4-, 5-, 6- and/or 7-¹⁵NO₂-*N*-AcTrp.

¹⁵N-CIDNP during nitration of *N*-AcTrp with H¹⁵NO₄

Peroxyxynitric acid is able to nitrate electron-rich aromatics at pH 2–3 [15]. During reaction of H¹⁵NO₄ with *N*-AcTrp at pH 2, the ¹⁵N-NMR signals of ¹⁵NO₃⁻ and of H¹⁵NO₄ appear in emission and in enhanced absorption, respectively. Emission is observed in additional signals (Figure 2(C), Table II), which might be due to 2-¹⁵NO₂-*N*-AcTrp (δ = -7 ppm), to 5-, 6- and 7-¹⁵NO₂-*N*-AcTrp (δ = 3 ppm) and to 4-¹⁵NO₂-*N*-AcTrp (δ = 7 ppm), in analogy to the reaction of *N*-AcTrp with H¹⁵NO₃. The signals at δ ~ 25 ppm point to some unidentified products. The effects in the ¹⁵N-NMR signals of ¹⁵NO₃⁻ and H¹⁵NO₄ are explained by reactions of H¹⁵NO₄ with H¹⁵NO₂ and by decay of H¹⁵NO₄ [15], those in ¹⁵NO₂-*N*-AcTrp by reactions of *N*-AcTrp^{•+} and ¹⁵NO₂ in radical pairs [*N*-AcTrp^{•+}, ¹⁵NO₂]^F, see Scheme 5. Radical cations of *N*-AcTrp are formed by oxidation with H¹⁵NO₄ or some RNS like ¹⁵NO⁺, ¹⁵N₂O₃, or ¹⁵NO₃ which are present during the decay of H¹⁵NO₄.

¹⁵N-CIDNP during nitration of Trp with preformed and *in situ* generated peroxyxynitrite-¹⁵N

Reactions of peroxyxynitrite with Trp and *N*-AcTrp have thoroughly been studied [5,22,23], see Table I. The most important reaction is nitration at the 6-position of the indole system leading to 6-NO₂-Trp or 6-NO₂-*N*-AcTrp. Nitrations at the ring positions 1, 4, 5 and 7 are also reported. Besides, a few additional products are formed which have not been listed in Table I and will not be discussed in the following. For taking ¹⁵N-NMR spectra at pH 7, the reaction of

peroxyxynitrite-¹⁵N with Trp was performed at 268 K, because CIDNP effects can only be observed, if reaction times are similar to nuclear relaxation times which are in the order of minutes [33]. As the protonated form of peroxyxynitrite decays with a rate of 1.3 s⁻¹ at 298 K [34], it is impossible to observe ¹⁵N-CIDNP at physiological pH values and room temperature with preformed peroxyxynitrite. At 268 K and pH 7.5, the decay rate of peroxyxynitrite is about 0.0025 s⁻¹ [35] giving reaction times of 10 min [10]. During the reaction, ¹⁵N-NMR signals of ¹⁵NO₂⁻ and ¹⁵NO₃⁻ appear within 3 min indicating the end of the reaction at that time. ¹⁵N-CIDNP effects are not observed. Details of the reaction conditions are given in Table IV. The results agree with those reported during the peroxyxynitrite-¹⁵N decay in the absence of Trp [9,10]. At peroxyxynitrite concentrations of 0.03 M, the reaction of peroxyxynitrite with Trp is slower than the induced decomposition reaction of peroxyxynitrite taking place under the given conditions [10].

The reported reactions of peroxyxynitrite with Trp have been performed at peroxyxynitrite concentrations of about 1 mM which are too small for CIDNP experiments because of the low sensitivity of ¹⁵N-NMR spectroscopy. This difficulty was circumvented by generating peroxyxynitrite-¹⁵N *in situ* using the Baeyer-Villiger reaction [1], see Figure 3 and Table IV. At pH 5, it is finished after about 10 min using [Na¹⁵NO₂] = 0.05 M and [H₂O₂] = 1 M.

A ¹⁵N-NMR spectrum of a solution of Na¹⁵NO₂ and Trp in H₂O/D₂O in the presence of NaHCO₃ is given in Figure 3(A). The signal of Na¹⁵NO₂ at δ = 242 ppm is not shown in Figure 3. After adding H₂O₂ to the solution, strong absorption and emission

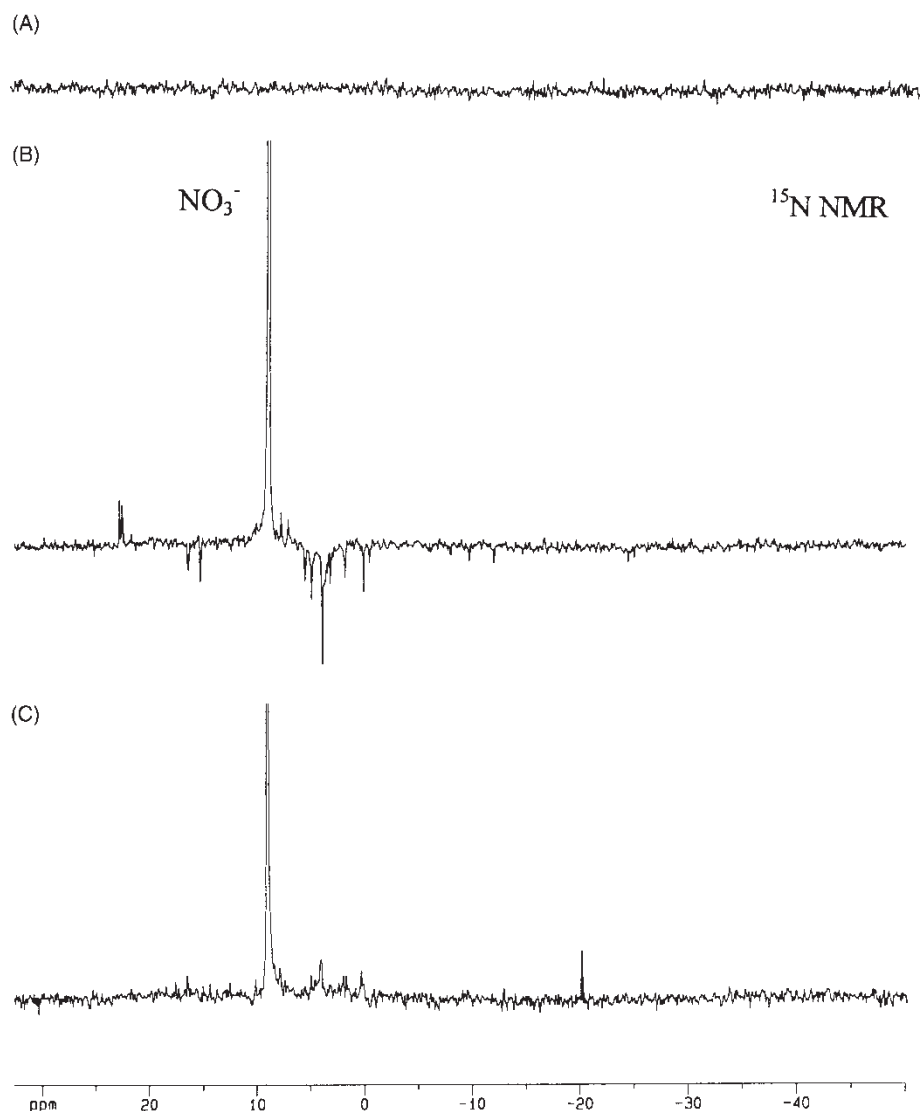
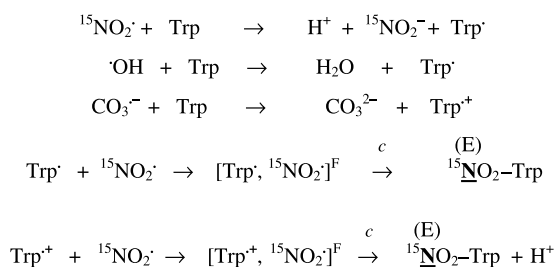


Figure 3. ^{15}N -NMR spectra of $\text{Na}^{15}\text{NO}_2$, NaHCO_3 and Trp at pH 5 in $\text{H}_2\text{O}/\text{D}_2\text{O}$ (A) before adding H_2O_2 (1 scan), (B) 2 min after adding H_2O_2 (1 scan), (C) 100 min later (900 scans).

signals are observed (Figure 3(B), Table IV). After reaction, the spectrum given in Figure 3(C) was taken using 900 scans. Besides the ^{15}N -NMR signals at $\delta = 9$ ppm and $\delta = -20$ ppm due to $^{15}\text{NO}_3^-$ and H^{15}NO_4 , it shows signals at $\delta = 1$ ppm and $\delta = 4$ ppm which are assigned to nitration products of Trp. The

assignment is supported by chemical shift calculations of nitrated 3-methylindole (Table III), and the assignment during reaction of Trp with H^{15}NO_3 , see above. The signal at $\delta = 4$ ppm is then due to 4- $^{15}\text{NO}_2$ -Trp, the signal at $\delta = 1$ ppm to 5-, 6- and 7- $^{15}\text{NO}_2$ -Trp. Signals which might be caused by 1-, 2- and 3- $^{15}\text{NO}_2$ -Trp are not detected.

During the reaction, the signal of $^{15}\text{NO}_3^-$ at $\delta = 8$ ppm shows enhanced absorption. Additionally, the ^{15}N -NMR signal of $^{15}\text{NO}_2^-$ at $\delta = 242$ ppm which is not given in Figure 3 shows emission. The ^{15}N -CIDNP effects are also observed in the absence of Trp and are generated in radical pairs $[\text{CO}_3^{\cdot-}, ^{15}\text{NO}_2]^{\text{S}}$ formed during decay of $\text{O}=\text{H}^{15}\text{NO}-\text{OCO}_2^-$ [10], see Scheme 2. H^{15}NO_4 ($\delta = -20$ ppm) is also formed in the absence of Trp [19]. The emission signals between $\delta = 20$ ppm and $\delta = -15$ ppm are caused by additional reaction products. The intensive emission signals at $\delta = 4$ ppm and $\delta = 1$ ppm are due to



Scheme 6. ^{15}N -CIDNP during reaction of peroxyntirite- ^{15}N with Trp and Trp derivatives.

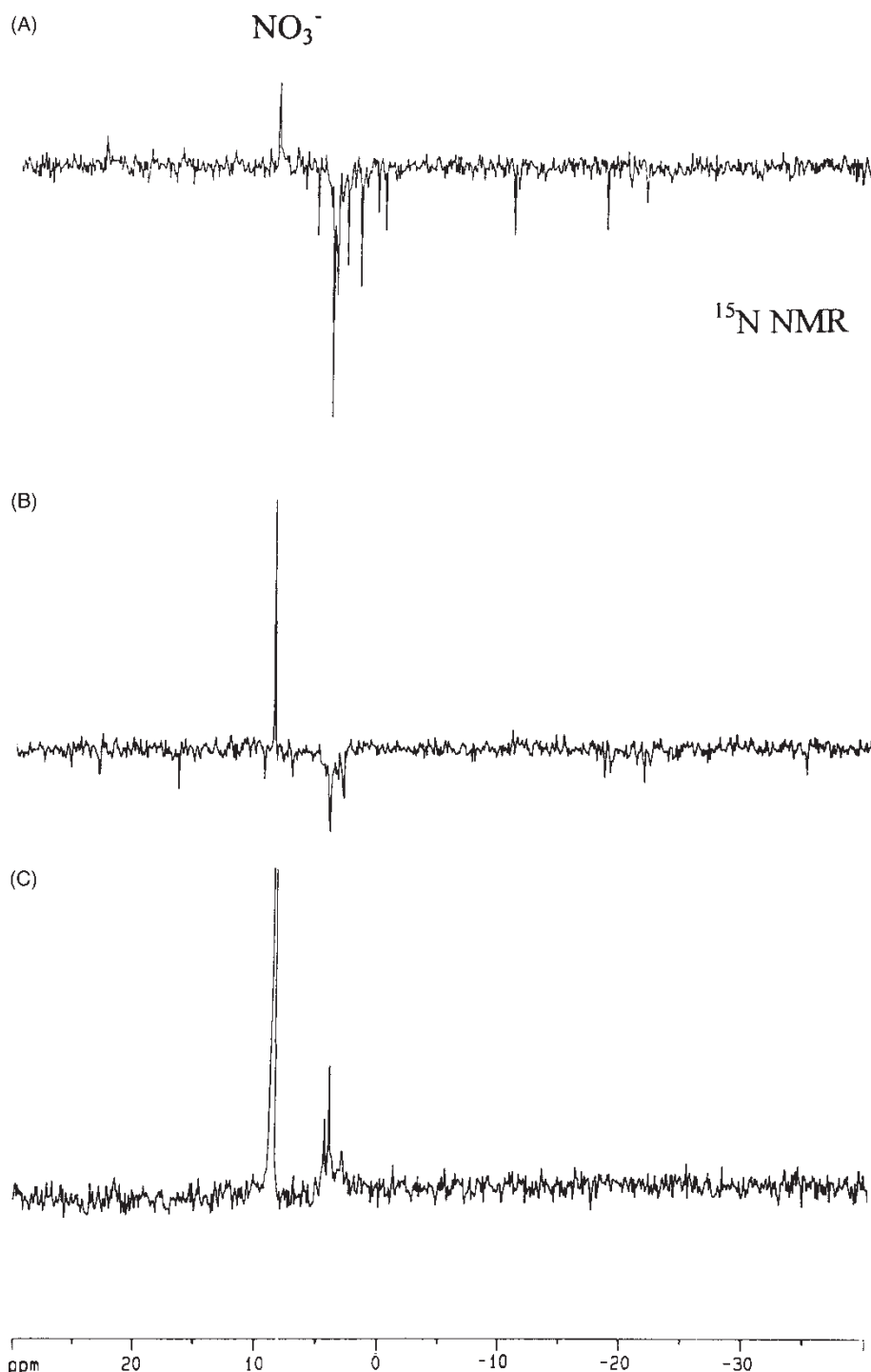


Figure 4. ^{15}N -NMR spectra of *N*-AcTrp in $\text{H}_2\text{O}/\text{D}_2\text{O}$ (A) during reaction of peroxynitrite- ^{15}N at pH 5 2 min after mixing the reactants (1 scan), (B) during reaction with H_2O_2 , $\text{Na}^{15}\text{NO}_2$ and HRP at pH 7 2 min after mixing the reactants (20 scans, delay time 20 s), (C) 1000 min later (7421 scans).

4- $^{15}\text{NO}_2$ -Trp and 5-, 6- and/or 7- $^{15}\text{NO}_2$ -Trp. The signals at $\delta = 15$ ppm and $\delta = 16$ ppm are assigned to the unstable *ipso* substitution products 3- $^{15}\text{NO}_2$ -Trp. One of the weak signals around $\delta = -10$ ppm might be due to 2- $^{15}\text{NO}_2$ -Trp. A signal at $\delta \sim -45$ ppm which might point to the formation of 1- $^{15}\text{NO}_2$ -Trp is not observed. Two enhanced

absorption signals at $\delta \sim 22$ ppm could not be assigned.

The occurrence of ^{15}N -CIDNP proves the radical mechanism of the $^{15}\text{NO}_2$ -Trp formation. Radicals Trp^\cdot or radical cations $\text{Trp}^{\cdot+}$ are generated by reaction of $^{15}\text{NO}_2$ or $\text{CO}_3^{\cdot-}$ with Trp [5,29], see Scheme 6. Polarized $^{15}\text{NO}_2$ -Trp is then formed by recombination

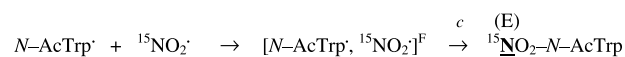
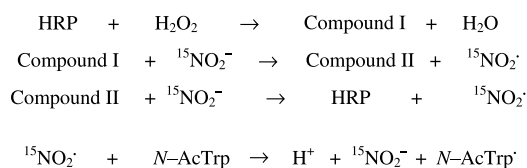
of Trp[•] or Trp^{•+} with ¹⁵NO₂⁻ in radical pairs [Trp[•], ¹⁵NO₂⁻]^F or [Trp^{•+}, ¹⁵NO₂⁻]^F.

Radicals ¹⁵NO₂⁻, OH[•], and CO₃^{•-} are formed during decay of peroxyxynitrite-¹⁵N and the peroxyxynitrite-¹⁵N-CO₂ adduct (Scheme 1) giving radicals Trp[•] or radical cations Trp^{•+}. The emission-type effect in the ¹⁵N-NMR signals of ¹⁵NO₂-Trp is explained using the observed *g* values of NO₂⁻ (2.000) and Trp[•] (2.003) [29,30], see Scheme 2. The reported p*K* value of 4.3 for Trp[•] indicates that Trp[•] and Trp^{•+} should be present under the reaction conditions (pH 5) [31]. As Trp[•] and Trp^{•+} should have similar spin densities at the indole ring positions [32], see Figure 1, the product yields of the different Trp nitration products should be similar, too, and do not allow distinction between Trp[•] and Trp^{•+} as free radical intermediates.

¹⁵N-CIDNP experiments have also been performed in CO₂-free solutions during reaction of peroxyxynitrite with *N*-AcTrp, see Figure 4(A) and Table IV. The ¹⁵N-CIDNP pattern looks similarly as that observed in the presence of CO₂. It is explained in an analogous manner as described (Scheme 4). The enhanced absorption of the ¹⁵N-NMR signal due to ¹⁵NO₃⁻ is missing (Scheme 2), and the ¹⁵N-NMR signal of H¹⁵NO₄ shows emission.

¹⁵N-CIDNP during nitration of *N*-AcTrp with the system H₂O₂/¹⁵NO₂⁻/HRP

It has been shown that ¹⁵N-CIDNP observed in acidic medium is caused by reactions of nitrogen dioxide with independently generated tryptophanyl radicals or radical cations in radical pairs [Trp[•], ¹⁵NO₂⁻]^F and/or [Trp^{•+}, ¹⁵NO₂⁻]^F. For confirming this at physiological pH values, radicals ¹⁵NO₂⁻ were generated from ¹⁵NO₂⁻ and H₂O₂ in the presence of HRP. It is well-known that Trp, *N*-AcTrp and *N*-AcTrp-NH₂ are nitrated by H₂O₂/NO₂⁻ in the presence of various peroxidases, see Table I [6,24,25]. Figure 4(B) shows a ¹⁵N-NMR spectrum taken during reaction of H₂O₂ with Na¹⁵NO₂⁻ at pH 7 in the presence of HRP. It also shows emission lines in the region between δ = -25 ppm and δ = 20 ppm indicating the formation of ¹⁵NO₂-*N*-AcTrp by recombination of ¹⁵NO₂⁻ with radicals *N*-AcTrp[•] during the reaction [36,37]. The most intensive signals between



Scheme 7. ¹⁵N-CIDNP during reaction of H₂O₂ and Na¹⁵NO₂ with *N*-AcTrp in the presence of HRP.

Table V. ¹⁵N-CIDNP during nitration of Mel with RNS at 298 K.

System	¹⁵ N-NMR signals	CIDNP*
Peroxyxynitrite- ¹⁵ N [†] , Mel (0.05 M), pH 5 (Figure 5b)	- 19 (H ¹⁵ NO ₄)	E
	6 (4- ¹⁵ NO ₂ -Mel)	E
	8 (¹⁵ NO ₃ ⁻)	N (A) [‡]
	16 (not assigned) [¶]	E
Peroxyxynitrite- ¹⁵ N [†] , Mel (0.05 M), NaHCO ₃ (0.05 M), pH 5 (Figure 5b)	- 19 (H ¹⁵ NO ₄)	E
	6 (4- ¹⁵ NO ₂ -Mel)	E
	8 (¹⁵ NO ₃ ⁻)	A
	22 (not assigned) [¶]	E
H ¹⁵ NO ₄ [§] , Mel (0.05 M), pH 2 (Figure 5c)	- 20 (H ¹⁵ NO ₄)	A
	6 (4- ¹⁵ NO ₂ -Mel)	E
	8 (¹⁵ NO ₃ ⁻)	E

* E: emission, A: enhanced absorption, N: no CIDNP; [†] Generated *in situ* by reaction of Na¹⁵NO₂ (0.05 M) with H₂O₂ (1 M); [‡] The signal is slightly enhanced because traces of CO₂ cause a decomposition of the peroxyxynitrite-¹⁵N in part via the CO₂ adduct; [¶] 3- or 5-¹⁵NO₂-Mel (see Table VI); [§] Generated *in situ* by reaction of Na¹⁵NO₂ (0.15 M) with H₂O₂ (1 M).

δ = 2 ppm and δ = 5 ppm might be assigned to 4-, 5-, 6- and 7-¹⁵NO₂-*N*-AcTrp again. They are also observed after reaction in a spectrum taken with 7421 scans (Figure 4(C)). A possible reaction mechanism is given in Scheme 7.

¹⁵N-CIDNP during reaction of Mel with peroxyxynitrite-¹⁵N and H¹⁵NO₄

Nitration of melatonin (Mel) by peroxyxynitrite leads to 4-nitromelatonin (4-NO₂-Mel) as the main product and 6-nitromelatonin (6-NO₂-Mel) as a side product [16]. In the following, ¹⁵N-CIDNP studies will be described using peroxyxynitrite-¹⁵N and H¹⁵NO₄ as nitrating agents.

During reaction of Mel with peroxyxynitrite-¹⁵N in the absence of NaHCO₃, a spectrum was taken shown in Figure 5(A) and described in Table V. An emission

Table VI. Quantum-chemically calculated isotropic absolute shielding constants and ¹⁵N chemical shifts (δ, in ppm against nitrobenzene-¹⁵N).

Molecule	Isotropic shielding constants*		Isotropic chemical shifts	
	B1LYP	B3LYP	B1LYP	B3LYP
Nitrobenzol	-125.2	-121.5	0.0	0.0
1- NO ₂ -2	-80.3	-76.2	-44.9	-45.3
2- NO ₂ -2	-112.9	-108.5	-12.3	-13.0
3- NO ₂ -2	-153.3	-149.8	28.1	28.3
4- NO ₂ -2	-132.8	-128.2	7.6	6.7
5- NO ₂ -2	-156.4	-153.3	31.2	31.8
6- NO ₂ -2	-129.3	-124.8	4.1	3.3
7- NO ₂ -2	-125.8	-121.2	0.6	-0.3

* Isotropic absolute shielding constants were calculated using the GIAO protocol at the DFT/aug-cc-pVDZ//DFT/aug-cc-pVDZ level of theory. During these calculations solvation corrections (CH₃CN for nitrobenzol, H₂O for all others) with the CPCMC solvation model were performed at the same level of theory [21].

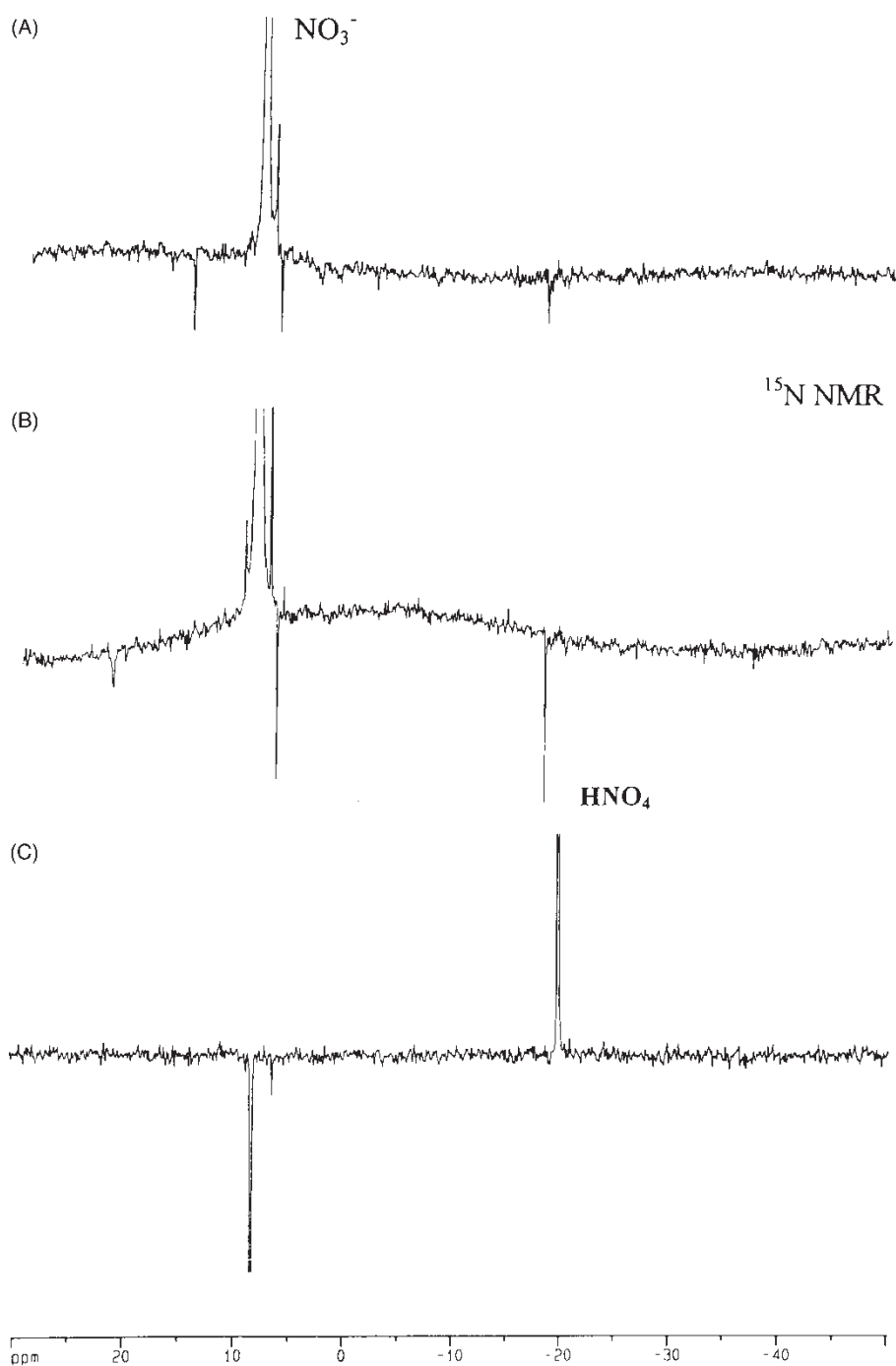
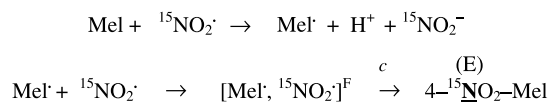


Figure 5. ^{15}N -NMR spectra of Mel in $\text{H}_2\text{O}/\text{D}_2\text{O}$ 2 min after mixing the reactants (20 scans, delay time 20 s) with (A) peroxynitrite- ^{15}N at pH 5, (B) peroxynitrite- ^{15}N with NaHCO_3 at pH 5, (C) H^{15}NO_4 at pH 2.

signal at $\delta = -19$ ppm is due to H^{15}NO_4 and observed in the absence of Mel, too. Additionally, two emission signals at $\delta = 6$ ppm and $\delta = 16$ ppm only appear in the presence of Mel. The first one is assigned to 4- $^{15}\text{NO}_2$ -Mel. For supporting this, a quantum-chemical calculation of ^{15}N chemical shifts has been performed with 3-methyl-5-methoxy-indane 2 as a model compound (Table VI), giving $\delta = 6.6$ ppm for 4- $^{15}\text{NO}_2$ -2. The second one was not assigned; it might be due to 3- or 5- $^{15}\text{NO}_2$ -Mel. In the presence of

NaHCO_3 , a similar spectrum was observed (Figure 5(A)). It does not show the signal at $\delta = 16$ ppm but another one at $\delta = 22$ ppm which was not assigned either. The ^{15}N -NMR signal of $^{15}\text{NO}_3^-$ appears in enhanced absorption as it was also observed in the absence of Mel.

During reaction of H^{15}NO_4 with Mel, the spectrum given in Figure 5(C) was taken showing the ^{15}N -NMR signals of $^{15}\text{NO}_3^-$ and of H^{15}NO_4 in enhanced absorption and emission. The signal at $\delta = 6$ ppm

Scheme 8. ^{15}N -CIDNP during nitration of melatonin with RNS.

shows emission, too, which supports its assignment to 4- $^{15}\text{NO}_2$ -Mel.

The ^{15}N -CIDNP effects in $^{15}\text{NO}_3^-$ and H^{15}NO_4 are explained as described, see Schemes 4 and 5. The emission-type effect in the ^{15}N -NMR signal of 4- $^{15}\text{NO}_2$ -Mel is explained in the same manner as those observed in $^{15}\text{NO}_2$ -Tyr and $^{15}\text{NO}_2$ -Trp, see Scheme 8.

Conclusions

The appearance of ^{15}N -CIDNP during reaction of Trp, *N*-AcTrp and Mel with various nitrating systems indicates the radical character of the nitration. Formation of 6- NO_2 -Trp might be used as an additional marker for the endogeneous intermediacy of RNS. Of course, the detection of endogenous 6- NO_2 -Trp is more difficult than the detection of endogenous 3- NO_2 -Tyr because of both the lower concentrations of Trp in proteins compared to Tyr and the well known electron transfer from tryptophanyl radicals to tyrosine [38] which should favor the formation of 3- NO_2 -Tyr at the expense of 6- NO_2 -Trp. Therefore, the presence of both 6- NO_2 -Trp and 3- NO_2 -Tyr would indicate the occurrence of a massive nitrative stress whereas the single detection of 3- NO_2 -Tyr indicated “only” the intermediacy of nitrogen dioxide [39].

The ^{15}N -CIDNP results do not rule out the possibility of non-radical nitrations. 4- NO_2 -Trp is formed as the only nitration product of Trp during reaction with a complex between *Deinococcus radiodurans* NOS and a tryptophanyl-tRNA synthetase [40]. The missing component of the nitration product 6- NO_2 -Trp makes a different nitration mechanism likely.

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